# Fosgonimeton, a Novel, Small-Molecule **Positive Modulator of** the HGF/MET System, Is Neuroprotective in Primary Neuron Culture

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## CONCLUSIONS

Fosgo-AM increased the number of neurons in culture and enhanced neurite outgrowth

Cortical neurons treated with fosgo-AM exhibited significant protection against neurological insults that are central to neurodegeneration such as mitochondrial dysfunction, excitotoxicity, inflammation, and oxidative stress

The major signaling pathways of HGF (via MET) mediate the neuroprotective effect of fosgonimeton against glutamate excitotoxicity

## **KEY TAKEAWAY**

Neuroprotective and neurotrophic effects of fosgo-AM, the active metabolite of fosgonimeton, highlight its therapeutic potential to restore neuronal health and slow or reverse neurodegeneration

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### Acknowledgments

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## Figure 1. Positive modulators of the HGF/MET pathway may counteract neurodegeneration through multiple mechanisms

- Fosgonimeton rapidly converts to a brain-penetrant active metabolite (fosgo-AM) after administration

### Network growth assay

- Primary cortical neurons from rat embryos at embryonic day 15 were cultured for 2 days before testing
- Cells were fixed and immunostained with a mouse monoclonal antibody against MAP2 to reveal neuronal structures, including neurites, and were counterstained with
- Hoechst dye to reveal nuclei

## Neuroprotection assay

- Primary cortical neurons from rat pups on postnatal days 1-3 were cultured for 35-40 days before testing • Neurons were treated with vehicle or fosgo-AM (1.0 nM, 10 nM, 100 nM, or 1000 nM) for 15 minutes, then given one of the following neurotoxic compounds: MPP<sup>+</sup>, glutamate, LPS, or  $H_2O_2$  for 24 hours
- Primary neuron cultures contain small amounts of HGF,<sup>4</sup> the activity of which fosgo-AM modulates
- The Cell Titer-Glo luminescent cell viability assay (Promega), which measures ATP levels, was used to calculate cell viability for each experimental group

## MET occlusion assay

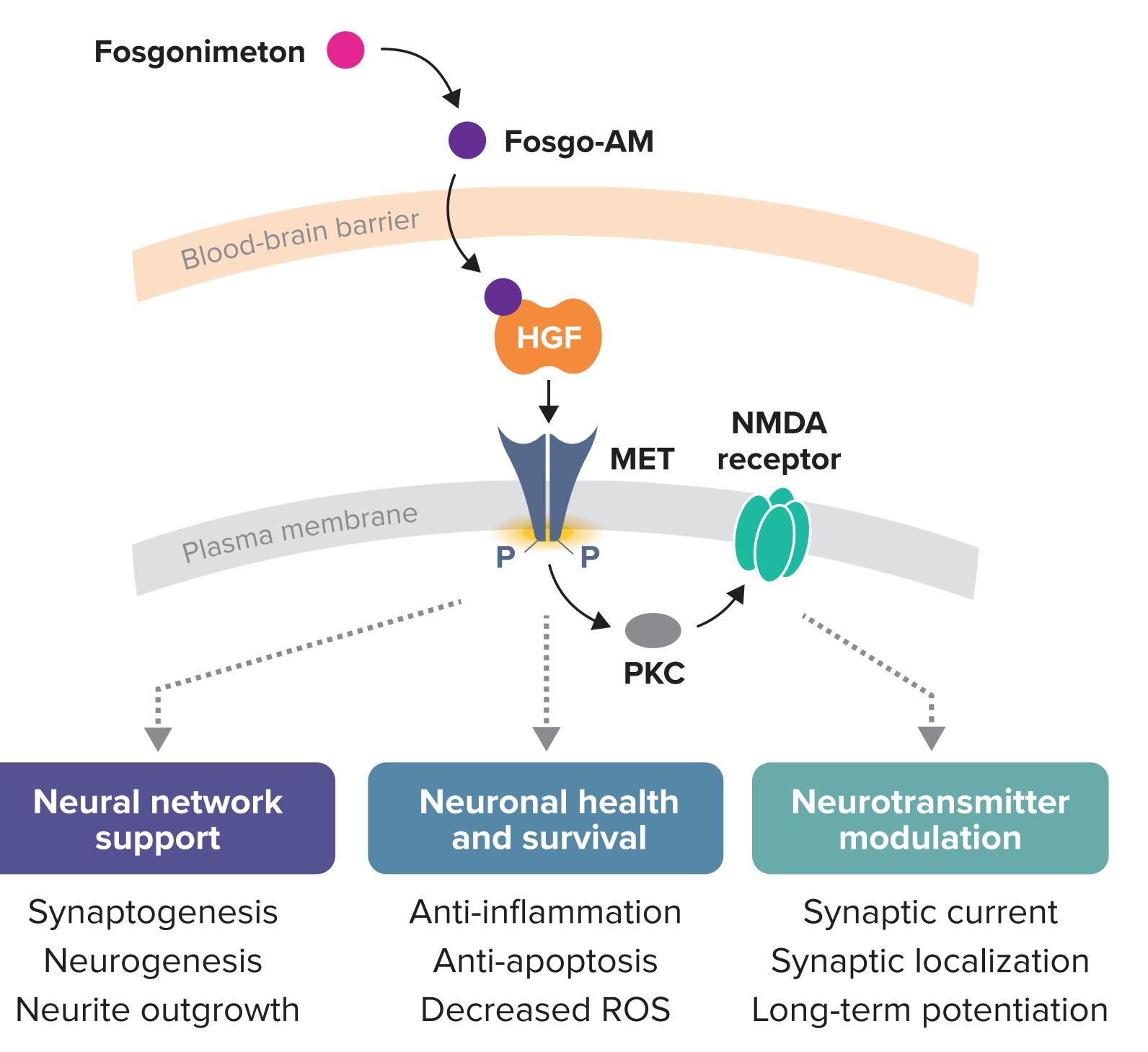
- Primary cortical neurons from rat embryos at embryonic day 15 were cultured for 12 days before testing
- Day 13 neuronal cultures were preincubated for 60 minutes with one of the following inhibitors of MET-associated molecules (or vehicle): GSK-690963 (100 nM; AKT inhibitor), calphostin C (200 nM; PKC inhibitor), KN-62 (2 μM; CamKII inhibitor), or PD98059 (10 μM; MEK inhibitor) (supplemental materials, QR code)
- cultures for the final 20 minutes of preincubation • At the end of the preincubation period, a neurotoxic insult with glutamate (20 µM) was applied for 20 minutes, in the presence of fosgo-AM/HGF and inhibitors • After the 20-minute injury with glutamate, test compounds and inhibitors were reintroduced for 24 hours

## Statistical analyses

- All data are displayed as mean + SEM
- EnVision instrument software (PerkinElmer)



## INTRODUCTION



HGF signaling through the MET receptor promotes neuronal growth and survival from prenatal development through adulthood<sup>1</sup>

• HGF/MET signaling also engages neuroprotective mechanisms that counteract several pathological hallmarks of neurodegeneration, such as excitotoxicity, neuroinflammation, and oxidative stress-induced damage<sup>1-3</sup>

Positive modulators of HGF/MET may provide therapeutic benefit to patients with Alzheimer's disease, Parkinson's disease, ALS, multiple sclerosis, cerebral ischemia, and other injuries/degenerative diseases of the nervous system<sup>1</sup>

Fosgonimeton is a small-molecule positive modulator of HGF/MET being evaluated for use in the treatment of Alzheimer's disease (ACT-AD, NCT04491006; LIFT-AD, NCT04488419), as well as for Parkinson's disease dementia or dementia with Lewy bodies (SHAPE, NCT04831281)

## OBJECTIVE

To assess the neuroprotective effects of fosgo-AM in vitro on cultured rat primary cortical neurons

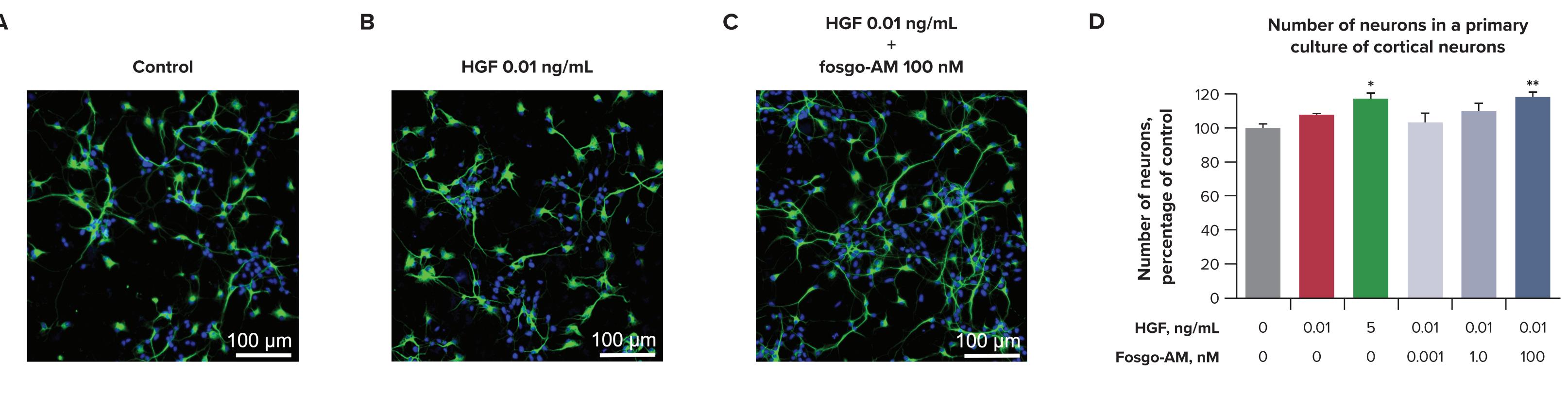
## METHODS

- A subthreshold concentration of HGF (0.01 ng/mL) with or without fosgo-AM (1 pM, 1 nM, or 100 nM) was added to neuronal cultures for 2 days
- The number of neurons and total length of neurite networks were automatically quantified with ImageXpress acquisition and MetaXpress analysis software (Molecular Devices)

- A subthreshold concentration of HGF (0.05 ng/mL), with or without fosgo-AM (100 nM), or an active concentration of HGF (5 ng/mL) was then added to neuronal
- Cells were fixed, immunostained, and quantified as for the network growth assay

• Prism 9 (GraphPad) 1-way ANOVA; post hoc Dunnett's multiple comparison test for network growth assays; and the Tukey multiple comparison test for cell viability assay

All P values are nominal and have not been corrected for multiple comparisons



Data presented as mean + SEM. \**P* < 0.05; \*\**P* < 0.01 compared with control (grey).

Data presented as mean + SEM. Statistical significance was determined by 1-way ANOVA with the Tukey posttest. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001 compared with insult (red).

Treatment with fosgo-AM at all evaluated/tested concentrations protected neurons from damage induced by neurotoxic insults that mimic several aspects of neurodegeneration: mitochondrial dysfunction (MPP<sup>+</sup>), excitotoxicity (glutamate), neuroinflammation (LPS), and oxidative stress (H<sub>2</sub>O<sub>2</sub>) • Treatment with 1  $\mu$ M fosgo-AM rescued cell viability against neurological insults with the following percent recoveries: 84.9% with MPP<sup>+</sup>, 61.6% with glutamate, 94.0% with LPS, and 144% with H<sub>2</sub>O<sub>2</sub>

+ +

10 100 1000

## Glutamate 20 µM

## GSK-690963 100 nM

Number of MAP2-positive neurons in culture following a 20-minute incubation in glutamate (20 µM) in the presence of (A) GSK-690963, (B) PD98059, (C) KN-62, or (D) calphostin C. Data presented in panels A-C were generated from the same experiment (plate), data presented in panel D were generated from a separate experiment (plate). Data presented as mean + SEM. Statistical significance was determined by 1-way ANOVA with the Tukey posttest. \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001; \*\*\*\**P* < 0.0001 compared with insult (red).

# oxygen species.

References 1. Desole C et al. Front Cell Dev Biol. 2021;9:683609. 2. Nicoleau C et al. Stem Cells. 2009;27:408-419. 3. Ko KR et al. Sci Rep. 2018;8:8316. 4. Gutierrez H et al. Development. 2004;131:3717-3726.

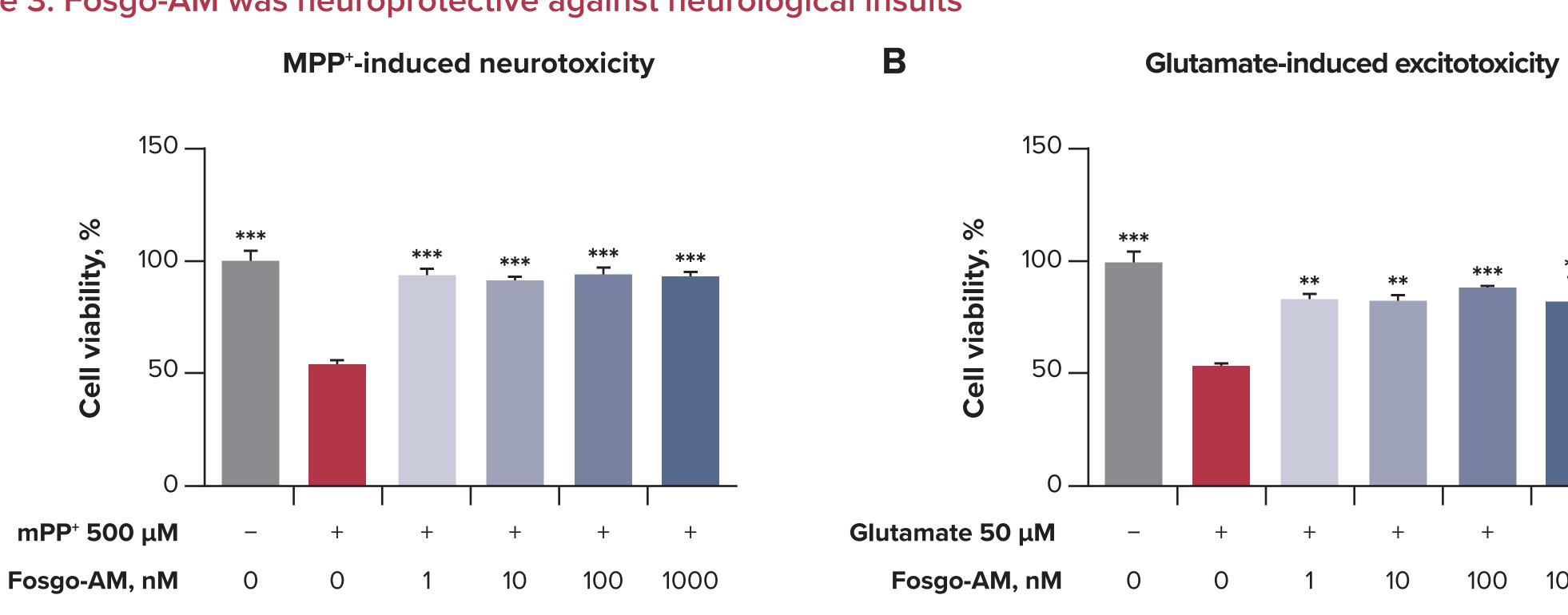
## RESULTS

### Figure 2. Fosgo-AM increased neuron numbers and enhanced neurite networks

Representative images from (A) control, (B) HGF 0.01 ng/mL, and (C) HGF 0.01 ng/mL + fosgo-AM 100 nM-treated primary cortical neurons and (E) total length of neurite network present after a 48-hour incubation period.

Statistical significance was determined by 1-way ANOVA with the Dunnett posttest.

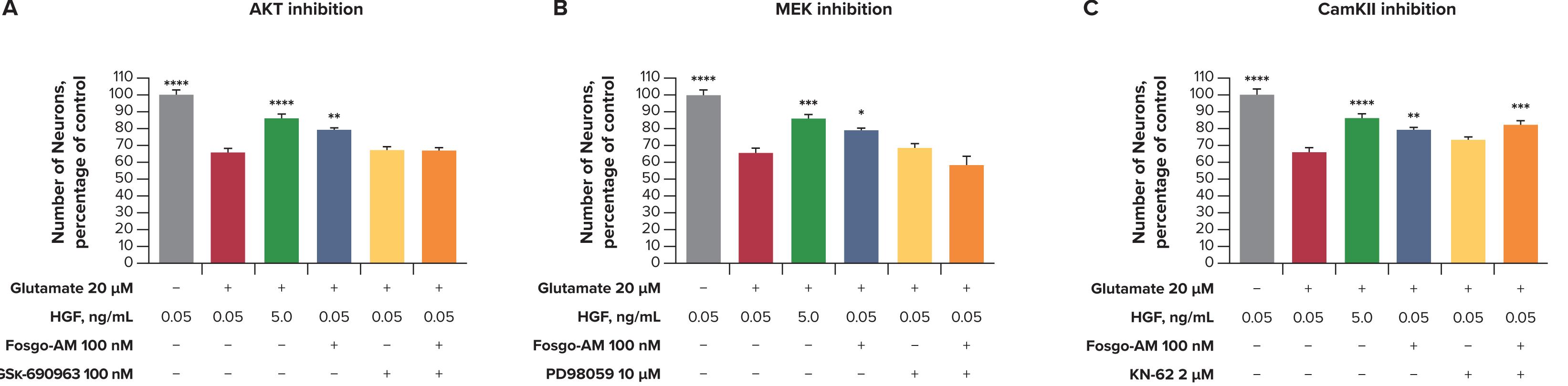
• Under basal conditions, the number of MAP2-positive neurons and total length of neurite network present after 48-hours incubation was significantly increased with 5 ng/mL HGF as well as with fosgo-AM (100 nM) + HGF (0.01 ng/mL), whereas HGF alone at 0.01 ng/mL did not have a significant effect - Fosgo-AM also augmented synaptic strength based on synaptobrevin-II staining intensity as an indication of the number of vesicles present at synapses (supplemental materials, QR code)



### Figure 3. Fosgo-AM was neuroprotective against neurological insults

Percentage of cortical neurons surviving in culture after exposure to (A) MPP<sup>+</sup>, (B) glutamate, (C) LPS, and (D)  $H_2O_2$ .

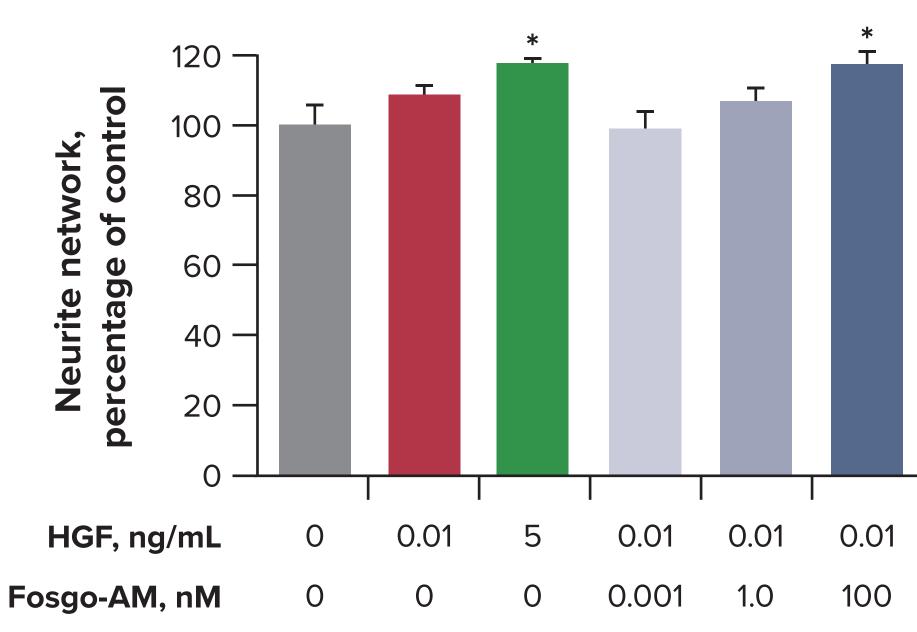
## Figure 4. Inhibition of the AKT, MEK, and PKC pathways downstream of HGF/MET signaling abolished the neuroprotective effects of fosgo-AM

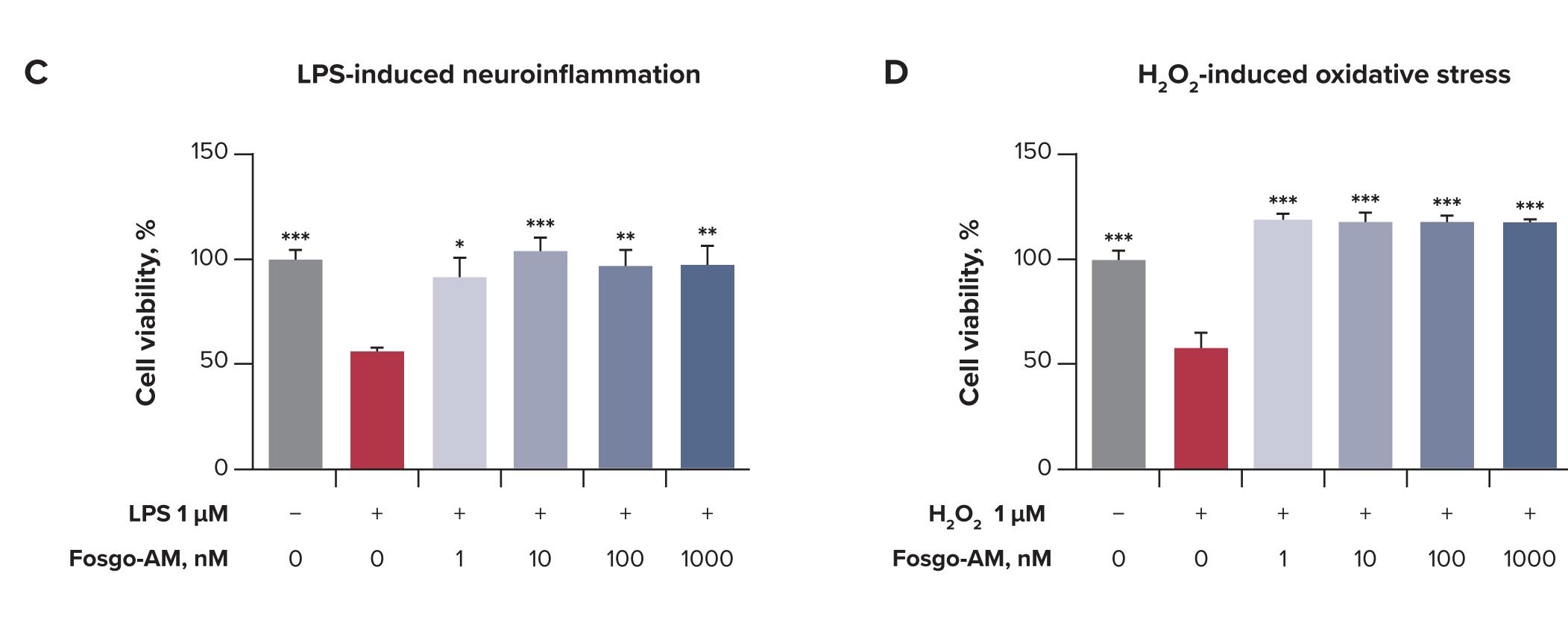


• Glutamate (20 µM for 20 minutes) induced a large and significant neuronal loss (neurons and neurite network). Fosgo-AM (100 nM) added 20 minutes before glutamate was able to significantly protect the neurons from the injury • Neuroprotective effects of fosgo-AM were fully abolished by GSK-690963 (AKT inhibition), calphostin C (PKC inhibition), and PD98059 (MEK inhibition) (supplemental materials, QR code) • In the presence of KN-62 (CamKII inhibition), fosgo-AM was still neuroprotective

s AKT, protein kinase B; ALS, amyotrophic lateral sclerosis; ANOVA, analysis of variance; ATP, adenosine triphosphate; CamKII, calcium/calmodulin-dependent protein kinase II; fosgo-AM, active metabolite of fosgonimeton; H<sub>2</sub>O<sub>2</sub>, hydrogen peroxide; HGF, hepatocyte growth factor; HTRF, homogeneous time-resolved fluorescence; LPS, lipopolysaccharide; MAP2, microtubule-associated protein kinase C; ROS, reactive structure in the structure

Neurite network length in a primary culture of cortical neurons





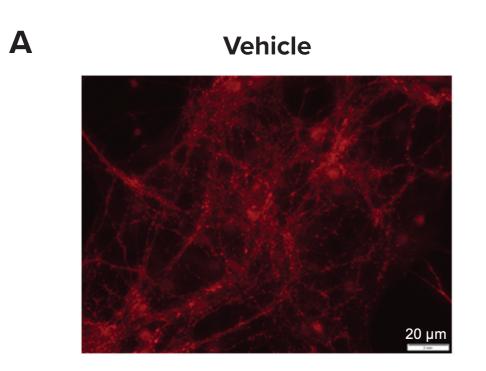
**PKC** inhibition D 90 – Glutamate 20 µM + + + +**HGF, ng/mL** 0.05 0.05 5.0 0.05 0.05 0.05 Fosgo-AM 100 nM - - + - + Calphostin C 200 nM – – – – – – + +

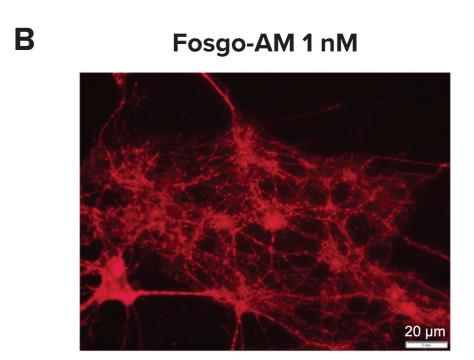
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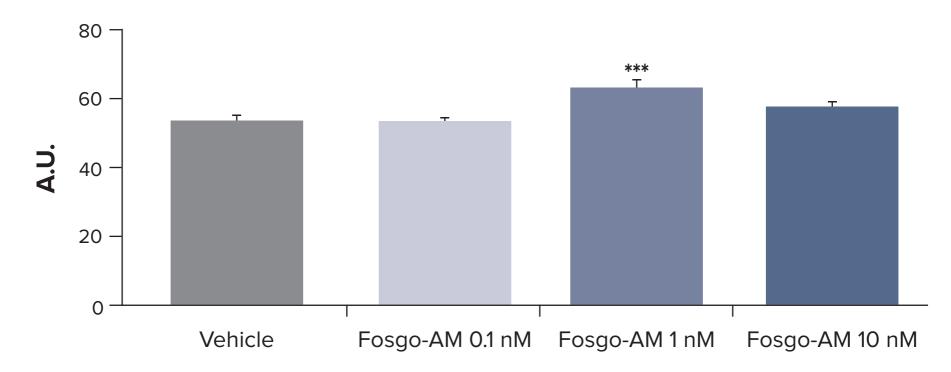
## **SUPPLEMENTAL INFORMATION**

Figure S1. Fosgo-AM increases synaptic strength based on synaptic vesicle density









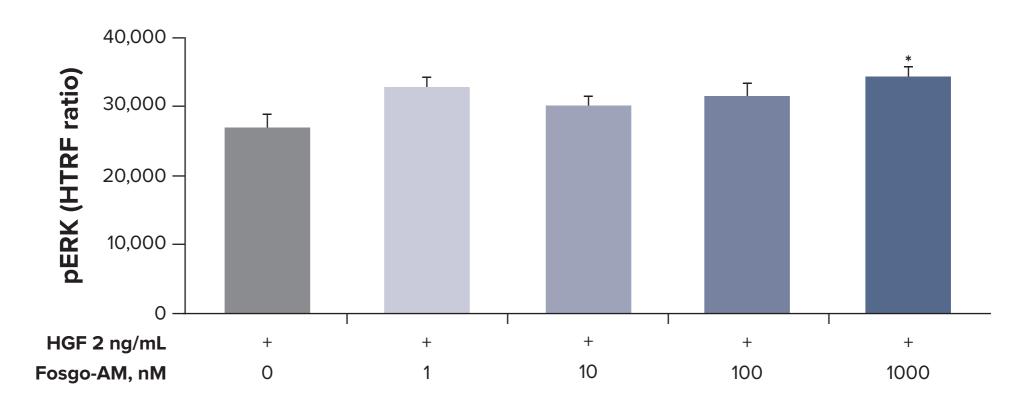
Primary rat hippocampal neurons were cultured and treated with fosgo-AM for 6 days and stained for synaptobrevin II, a marker of synaptic vesicles. Representative images from (**A**) vehicle and (**B**) fosgo-AM 1 nM highlight the effect of fosgo-AM on synaptic strength (relative abundance of presynaptic vesicles per synapse as measured by synaptobrevin II fluorescence intensity). (**C**) Fosgo-AM 1 nM significantly enhanced synaptic strength.

Data presented as mean + SEM.

Statistical significance was determined by 1-way ANOVA with the Dunnett posttest.

\*\*\*P < 0.001 compared with vehicle.

### Figure S2. Fosgo-AM enhances ERK phosphorylation

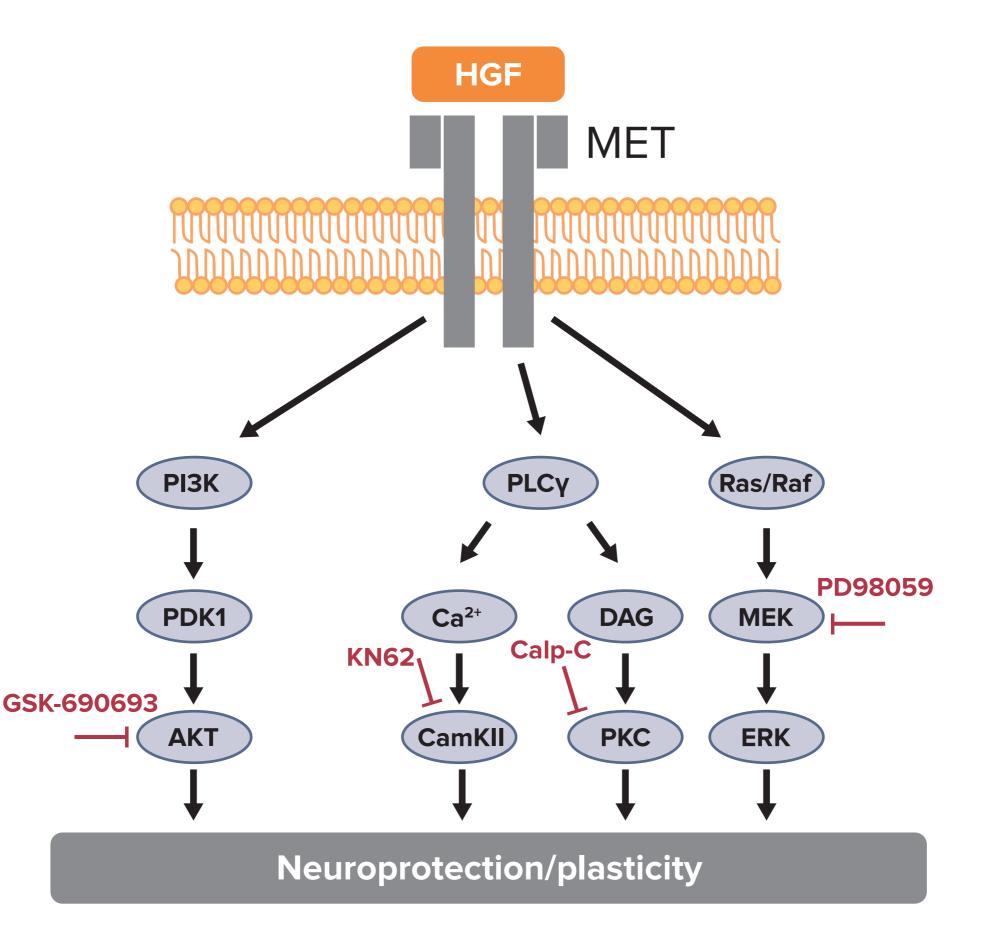


HEK-293 cells were subjected to HTRF with HGF (2 ng/mL) alone or with fosgo-AM (1 nM, 10 nM, 100 nM, or 1  $\mu$ M). pERK was measured using the advanced phospho-ERK (Thr202/Tyr204) cellular kit (#64AERPEG; CisBio). Treatment with fosgo-AM + HGF in cultured HEK-293 cells led to increased pERK relative to HGF alone (which does not significantly increase pERK at 2 ng/mL; data not shown) Data presented as mean + SEM.

Statistical significance was determined by 1-way ANOVA with the Dunnett posttest.

\*P < 0.05 compared with control (grey)

Figure S3. Inhibition of the AKT, CamKII, PKC, and MEK pathways downstream of HGF/MET signaling suggests potential mechanisms for fosgo-AM's neuroprotective effects



Inhibition with GSK-690693, KN62, Calp-C, or PD98059 (red) disrupts downstream elements of the HGF/MET pathway. While inhibition of AKT, PKC, and MEK resulted in disruption of fosgo-AM's neuroprotective effects, inhibition of CamKII did not.

Abbreviations AKT, protein kinase B; ALS, amyotrophic lateral sclerosis; Calp-C, calphostin C; CamKII, calcium/calmodulin-dependent protein kinase II; DAG, diacylglycerol; ERK, extracellular signal-regulated kinase; fosgo-AM, active metabolite of fosgonimeton; HEK-293, human embryonic kidney 293; HGF, hepatocyte growth factor; HTRF, homogeneous time-resolved fluorescence; MEK, mitogen activated protein kinase; MET, mitogen-activated protein kinase kinase; PDK1, phosphoinositide-dependent kinase 1; pERK, phosphorylation of ERK; PI3K, phosphatidyl-inositol 3 kinase; PKC, protein kinase C; PLCγ, phospholipase Cγ.

### Acknowledgments

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